**Storage conditions and stability**

STORE TRANSFIX® AT ROOM TEMPERATURE. Contents of unopened bottles are stable until the expiration date printed on the label. Contents of unopened bottles are stable for 1 month after opening.

**Evidence of deterioration**

The normal appearance of TransFix® is that of a clear green liquid. Any change in the physical appearance of the reagent may indicate deterioration and the reagent should not be used.

**Materials required but not supplied**

1. Evacuated blood collection tubes with anticoagulant (K3EDTA, ACD, heparin).
2. Pipettors and plastic tips capable of delivering 200μl to 1000μl.
3. Plastic storage rack.
4. Labels to indicate the specimens, samples have had TransFix® reagent added.

**Anticoagulant**

In whole blood samples an EDTA salt (K3EDTA) is the anticoagulant of choice for TransFix®. However ACD or heparin may also be used.

**Procedure**

No preparation is needed.

1. Determine volume of blood in the blood collection tube.
   - Note: In the Vacutainer system a full blood collection tube will contain 4ml of blood.

2. Carefully remove the blood sample tube cap.

3. Pipette into the blood collection tube an appropriate volume of Transfix® at the specified ratio of 200μl Transfix® per 1000μl of blood.

4. Replace blood collection tube cap, ensuring no leakage is evident.

5. Hand mix by repeated inversion (5x) to distribute the Transfix® reagent completely through the blood sample.

6. Label blood collection tube to indicate that Transfix® reagent has been added.

7. Place samples in a rack for subsequent cellular antigen assay. TransFix® treated samples are stable for up to 10 days at 4°C. 7 days at room temperature and 3 days at 37°C.

**PLEASE NOTE:**

**BLOOD SAMPLES STORED AT 4°C MUST BE ALLOWED TO RETURN TO ROOM TEMPERATURE BEFORE ANY CELLULAR ANTIGEN ASSAYS ARE PERFORMED.**

Stored Transfix® treated samples must be mixed thoroughly by inversion (25x) to resuspend cells before analysis by flow cytometry. Cell counts from Transfix® treated samples must be divided by 0.83 to adjust for the dilution factor. Standard erythrocyte protocols may be used with Transfix® treated samples. Leukocyte populations should be identified using a CD45 fluorescence and orthogonal scatter, or equivalent gating.

**Limitations**

When using Transfix® with samples collected from leukemic patients it may be necessary to establish an appropriate volume of the reagent to add that is different to that quoted in the procedure. Volumes in the region of 0.5-1.0 ml per ml of blood may be more appropriate.

**Selected references**


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